

**{Exhibit 52}**

Boguslaski et al., UK Patent Application GB-2 045  
239 A, published October 29, 1980

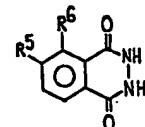
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**UK Patent Application (19) GB (11) 2 045 239 A**

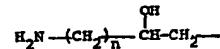
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Patents Act 1977.  
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(54) Intermediates useful in the  
synthesis of chemiluminescent -  
labeled conjugates for use in  
specific binding assays

(57) Novel compounds of the formula:



wherein one of R<sup>5</sup> and R<sup>6</sup> is hydrogen  
and the other is -NR'R"; R' is hydrogen  
or straight chain alkyl containing 1-4  
carbon atoms and R" is



wherein n=1 to 3, are intermediates in  
the synthesis of chemiluminescent -  
labeled conjugates useful in specific  
binding assays.

**GB 2 045 239 A**

**SPECIFICATION****Intermediates useful in the synthesis of chemiluminescent-labeled conjugates for use in 5 specific binding assays**

This invention relates to novel intermediates useful in the synthesis of chemiluminescent - labeled conjugates for use in specific binding assays for a 10 ligand, such as an antigen, hapten or antibody, in a liquid medium such as a body fluid. This Application is a divisional of our Application No. 7923660 (Serial No. 2 026 690) filed July 6th 1979.

The desirability of a convenient, reliable, and 15 non-hazardous means for detecting the presence of low concentrations of substances in liquids is self-evident. This is particularly true in the field of clinical chemistry where constituents of body fluids which may appear in concentrations as low as  $10^{-11}$  molar 20 are known to be of pathological significance. The difficulty of detecting such low concentrations is compounded in the field of clinical chemistry where sample size is usually quite limited.

Classically, substances have been detected in 25 liquids based on a reaction scheme wherein the substance to be detected is a necessary reactant. The presence of unknown is indicated by the appearance of a reaction product or the disappearance of a known reactant. In certain instances, such an assay 30 method may be quantitative, based on a measurement of either the rate of appearance of product or disappearance of reactant or measurement of the aggregate amount of product produced or reactant consumed in attaining equilibrium. Each assay reaction system is necessarily either limited to use in the 35 detection of only a small group of substances or is nonspecific.

The search for assay systems which are highly specific yet adaptable to the detection of a wide 40 range of substances has evolved the radioimmunoassay. In this system a known amount of a radiolabeled form of the substance to be detected is allowed to compete with the unknown for a limited quantity of antibody specific for the unknown. The 45 amount of the labeled form that becomes bound to antibody varies inversely with the level of unknown present. Inherent in the radio-immunoassay technique is the need to separate the labeled form of substance to be detected which becomes bound to antibody from that which does not become so bound. While various ways of accomplishing the required 50 separation have been developed, as exemplified in U.S. Patents Nos. 3,505,019; 3,555,143; 3,648,346; 3,720,760; and 3,793,445, all require at least one 55 separate manipulative step, such as filtering, centrifuging, or washing, to insure efficient separation of the bound-labeled form from the unbound-labeled form. The elimination of the separation step

would greatly simplify the assay and render it more useful to the clinical laboratory.

The use of radioactive materials in immunoassays has been eliminated to some degree by the use of enzyme-tagged materials in place of radiolabels. As exemplified by U.S. Patents Nos. 3,654,090 and 65 3,791,932, the manipulative steps necessary for carrying out the enzyme-tagged immunoassays are for the most part the same as those required in radioimmunoassays and include the cumbersome separation step. An additional disadvantage of using

70 enzyme-tagged materials is that each enzyme used as a tag must be individually chemically modified for use in the formation of the tagged conjugate. The use of other tagging materials has been suggested, such as the use of coenzymes or viruses, *Nature* 219:186 (1968) and the use of fluorescent labels, French Patent No. 2,217,350 corresponding to U.S. Patent No. 3,880,934.

While these radiolabeled and enzyme-tagged immunoassays may undergo future improvement in 75 terms of expansion of the range of substances detectable thereby or of simplification of the procedure, by their nature they will always require some type of separation step. Recently, a different approach was disclosed which does not require a separation step and therefore has been referred to as 80 a homogeneous system, in contrast to a heterogeneous system in which separation is essential. U.S. Patent No. 3,817,837 discloses a competitive binding assay method involving the steps of combining the liquid to be assayed with a soluble complex consisting of an enzyme as a labelling substance covalently bound to the ligand to be detected and with a soluble receptor, usually an antibody, for the ligand; and analyzing for the effect of the liquid 85 to be assayed on the enzymatic activity of the enzyme in the complex.

While this method has the advantage of not requiring a separation step because reaction between the 90 enzyme-bound-ligand complex and the receptor results in inhibition of the enzymatic activity of the enzyme in the complex, the method nonetheless is severely restricted in its ability to be adapted to widely varied assay requirements. For instance, it is clearly essential that in the fabrication of the 95 enzyme-bound-ligand complex, the substance or ligand to be detected must be coupled to the enzyme in a carefully controlled manner so that the coupling site is close to the enzymatically active site on the enzyme. This is required in order that upon reaction 100 between the complexed ligand and the receptor, the enzymatically active site is blocked. Enzymes vary greatly in their size, ranging in molecular weight from about 10,000 to 1,000,000. Thus, for a receptor in the form of an antibody having a molecular weight 105 of between 150,000 and 300,000 to be capable of physically blocking the active site on an average 110 between the complexed ligand and the receptor, the enzymatically active site is blocked. Enzymes vary greatly in their size, ranging in molecular weight from about 10,000 to 1,000,000. Thus, for a receptor in the form of an antibody having a molecular weight 115 of between 150,000 and 300,000 to be capable of physically blocking the active site on an average

Certain of the chemical formula(e) appearing in the printed specification were submitted in formal form after the date of filing.

The date of filing shown above is that provisionally accorded to the application in accordance with the provisions of Section 15(4) of the Patents Act 1977, and is subject to ratification or amendment at a later stage of the application proceedings.

enzyme of 500,000 molecular weight or greater, the coupling site must be precisely controlled. Due to the complex chemical structure of enzymes, precise control of such chemical linkage is indeed difficult, 5 and one would expect that even upon screening a wide variety of enzymes only a small number would be found to be of use in this homogeneous assay system.

Moreover, it is critical for the purpose of obtaining 10 quantitative test results to precisely control the ratio of the number of enzymes to the number of ligands in each enzyme - bound - ligand complex. Here also, the complex peptide structure of enzymes makes such control difficult. It would again be expected that 15 only a small number of enzymes would have suitable molecular structure to ensure necessary control of the ligand/enzyme ratio.

The prior art homogeneous assay method is stated to involve an enzyme amplification and thus 20 to be highly sensitive. However, since the labeling substance, namely the enzyme, is itself the limiting factor determining the sensitivity of the prior art assay method, the versatility of the method is severely restricted. The sensitivity is clearly limited 25 to the catalytic activity of the particular enzyme in the enzyme-bound-ligand conjugate. The versatility of the prior art method is therefore restricted not only by the coupling requirements for formation of a useful conjugate but also by the dependence of the 30 sensitivity of the assay that employs such conjugate on the activity of the particular conjugated enzyme.

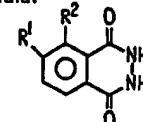
An additional disadvantage of the prior art homogeneous assay method arises in its application to the testing of biological fluids, such as urine and 35 serum. It is to be expected that significant amounts of the enzyme species comprised in the enzyme-bound-ligand conjugate may appear in the fluid sample to be tested thereby creating an uncontrollable background activity which would severely affect 40 the accuracy of the assay method. Therefore, in order to form an assay system that is useable in testing biological fluids of humans or animals, exotic enzymes not endogenous to such fluids must be selected for use in forming the enzyme-bound-ligand conjugate with the result that the versatility of 45 the assay method is even further restricted.

The present Invention provides novel Intermediates useful in the synthesis of chemiluminescent-labeled conjugates for use in specific binding assays 50 as described in our aforesaid Application No. 79.23660. (Serial No. 2 026 690)

In our Application No. 79.23660. (Serial No. 2 026 690), out of which this application has been divided, we have described and claimed a highly convenient, 55 versatile, and sensitive homogenous specific binding assay method and system based on the use of, as labeling substance, a reactant in a chemiluminescent reaction. The method is based, in part, on the fact that the reaction between a ligand and a specific binding partner thereof to one of which the said 60 reactant is coupled in the form of a conjugate, alters the activity of the reactant in the chemiluminescent reaction, which reaction thus serves as means for monitoring the specific binding reaction. Various 65 manipulative schemes involving various test com-

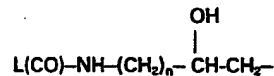
positions and devices may be employed in performing the method and are described in the aforesaid Application.

Particularly useful chemiluminescent-labeled conjugates described in our aforesaid Application are those of the formula:



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wherein one of R¹ and R², preferably R², is hydrogen and the other is -NR³R⁴; R³ is hydrogen or straight chain alkyl containing 1-4 carbon atoms, preferably ethyl, and R⁴ is



wherein n = 1 to 3, preferably 1, and L(CO)- is a specifically bindable ligand, or a binding analog thereof, bound through an amide bond.

The specifically bindable ligand or analog thereof in the present labeled conjugates, in terms of its chemical nature, usually is a protein, polypeptide,

90 peptide, carbohydrate, glycoprotein, steroid, or other organic molecule for which a specific binding partner is obtainable. In functional terms, the ligand will usually be an antigen or an antibody thereto; a hapten or an antibody thereto; or a hormone, vitamin, or drug, or a receptor or binding substance therefor. Most commonly, the ligand is an immunologically-active polypeptide or protein of

95 molecular weight between 1,000 and 4,000,000 such as an antigenic polypeptide or protein or an antibody; or is a hapten of molecular weight between 100 and 1,500.

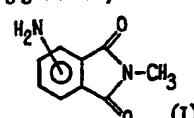
The aforesaid labeled conjugates are prepared usually by forming a peptide or amide couple between (1) an amino derivative of a chemiluminescent 105 aminophthalhydrazide (e.g., luminol or isoluminol) and (2) either the ligand, where such contains a carboxylic acid function, or a binding analog of the ligand (e.g., a derivative of the ligand) which analog contains the desired carboxylic acid function. Such

110 condensation reactions can be accomplished by reacting the amino derivative of the label directly with the carboxylic acid-containing ligand or ligand analog using conventional peptide condensation reactions such as the carbodiimide reaction [Science

115 144:1344 (1964)], the mixed anhydride reaction [Erlanger et al, *Methods In Immunology and Immuno-chemistry*, ed. Williams and Chase, Academic Press (New York 1967) p. 149], and the

120 acid azide and active ester reactions [Kopple, *Pep-tides and Amino Acids*, W. A. Benjamin, Inc. (New York 1966)]. See also for a general review Clin. Chem. 22:726 (1976).

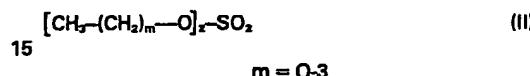
125 Preferably, however, the aforesaid chemiluminescent-labeled conjugates are prepared according to the following general synthetic sequence:



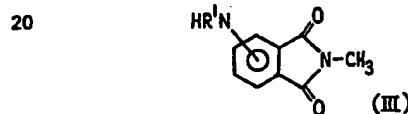
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The starting material for the synthesis is 3- or 4-amino-N-methylphthalimide (I) with the 3-amino compound [Wang et al, JACS 72:4887 (1950) and Flitsch, Chem. Ber. 94:2494 (1961)] to be used to 5 prepare luminol based labeled-conjugates and the 4-amino compound [Flitsch, Chem. Ber. 94:2494 (1961)] to be used to prepare isoluminol based labeled-conjugates.

Alkylation of the amino group in the phthalimide 10 (I) is obtained by reaction with a dialkyl sulfate (II) [Rodd, *Chemistry of Carbon Compounds*, vol. 1, Elsevier Publ. Co. (New York 1951) p. 337].

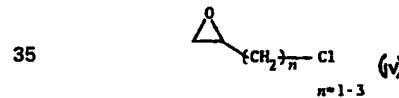


to yield the N-alkylated derivative (III)

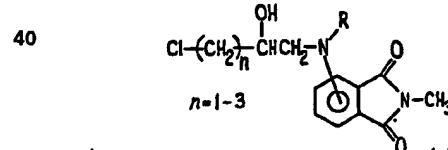


25 wherein R' is straight chain alkyl containing 1-4 carbon atoms.

Treatment of the phthalimide (I) or its N-alkylated derivative (III) with a chloro-epoxide (IV) [available from Aldrich Chemical Co., Milwaukee, Wisconsin 30 USA, or see Paul et al, Bull. Soc. Chim. Fr. 197 (1948) or Reppe et al, Justus Liebig's Annalen der Chemie 596:80-158 (1955)]

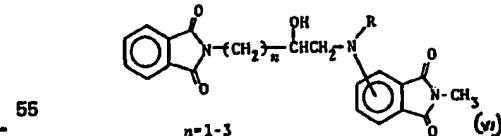


produces the chloro-intermediate (V)

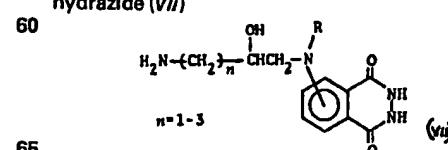


45 wherein R is hydrogen or straight chain alkyl containing 1-4 carbon atoms.

Reaction of the chloro-intermediate (V) with potassium phthalimide produces the *bis*-phthalimide intermediate (VI)



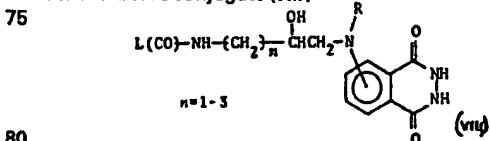
wherein R is the same as defined above, which upon treatment with hydrazine produces the amino-hydrazide (VII)



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wherein R again is the same as defined above.

Condensation of the amino-hydrazide (VII) with (a) the ligand to be labeled, where such contains a carboxylic acid function, (b) a binding analog of the ligand, such analog being a carboxylic acid derivative of the ligand, or (c) the ligand or an appropriate derivative of the ligand in the presence of a bifunctional coupling agent, produces the chemiluminescent-labeled conjugate (VIII)

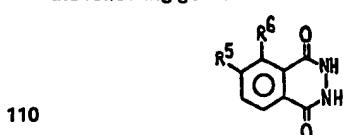


80 wherein R is the same as defined above and L(CO) represents the specifically bindable ligand, or a binding analog thereof (formed by derivation of the ligand and/or insertion of a bridge by a bifunctional coupling agent), bound through an amide bond.

Other variations of labeled conjugates based on the above-described synthetic scheme are clearly evident. In particular, various ring-substituted amino-N-methylphthalimides may be used as starting

90 material to produce ring-substituted labeled conjugates possessing substantially the same qualitative properties as the conjugates prepared according to the above-described scheme. Such conjugates will be recognized as equivalents and are exemplified by 95 the addition of one, two or more simple substituents to an available aromatic ring site, such substituents including without limitation, alkyl, e.g., methyl, ethyl and butyl; halo, e.g., chloro and bromo; nitro; hydroxyl; alkoxy, e.g., methoxy and ethoxy, and so forth.

100 As illustrated in the above-described synthetic scheme, the novel intermediate compounds of the present invention produced in the course of preparing the chemiluminescent-labeled conjugates have 105 the following general formula:



wherein one of R<sup>5</sup> and R<sup>6</sup>, preferably R<sup>6</sup>, is hydrogen and the other is -NR<sup>7</sup>R<sup>8</sup>; R<sup>7</sup> is hydrogen or straight chain alkyl containing 1-4 carbon atoms, preferably 115 ethyl, and R<sup>8</sup> is



wherein n = 1 to 3, preferably 1.

As stated hereinabove, the ligand which is comprised in the labeled conjugate or whose binding analog is comprised in the labeled conjugate is in 125 most circumstances an immunologically-active polypeptide or protein of molecular weight between 1,000 and 4,000,000 such as an antigenic polypeptide or protein or an antibody; or is a hapten of molecular weight between 100 and 1,500. Following will now 130 be presented various methods for coupling such

**Ligands or analogs thereof to the amino-derivative (VII) of the label through an amide bond.**

**Polypeptides and Proteins**

5 Representative of specifically bindable protein ligands are antibodies in general, particularly those of the IgG, IgE, IgM and IgA classes, for example hepatitis B antibodies; and antigenic proteins such as insulin, chorionic gonadotropin (e.g., HCG), carcinoma-embryonic antigen (CEA), myoglobin, hemoglobin, follicle stimulating hormone, human growth hormone, thyroid stimulating hormone (TSH), human placental lactogen, thyroxine binding globulin (TBG), intrinsic factor, trans-15 cobalamin, enzymes such as alkaline phosphatase and lactic dehydrogenase, and hepatitis-associated antigens such as hepatitis B surface antigen (HB<sub>s</sub>Ag), hepatitis B antigen (HB<sub>e</sub>Ag) and hepatitis core antigen (HB<sub>c</sub>Ag). Representative of polypeptide ligands 20 are angiotensin I and II, C-peptide, oxytocin, vasopressin, neuropephsin, gastrin, secretin, and glucagon. Since, as peptides, ligands of this general category possess numerous available carboxylic acid and amino groups, coupling to the amino-derivative of 25 the chemiluminescent label can proceed according to conventional peptide condensation reactions such as the carbodiimide reaction, the mixed anhydride reaction, and so forth as described hereinabove, or by the use of conventional bifunctional reagents 30 capable of coupling carboxylic acid or amino functions to the amino group in the label derivative as likewise described above. General references concerning the coupling of proteins to primary amines or carboxylic acids are mentioned in detail above.

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**Haptens**

Haptens, as a class, offer a wide variety of organic substances which evoke an immunochemical response in a host animal only when injected in the 40 form of an immunogen conjugate comprising the hapten coupled to a carrier molecule, almost always a protein such as albumin. The coupling reactions for forming the immunogen conjugates are well developed in the art and in general comprise the 45 coupling of a carboxylic acid ligand or a carboxylic acid derivative of the ligand to available amino groups on the protein carrier by formation of an amide bond. Such well known coupling reactions are directly analogous to the present formation of 50 labeled conjugates by coupling carboxylic acid ligands or binding analogs to the amino-derivative of the chemiluminescent label.

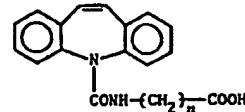
Hapten ligands which themselves contain carboxylic acid functions, and which thereby can be coupled directly to the amino-derivative of the label, include the iodothyronine hormones such as thyroxine and triiodothyronine, as well as other materials such as biotin, valproic acid, folic acid and certain prostaglandins. Following are representative synthetic 55 routes for preparing carboxylic acid binding analogs of hapten ligands which themselves do not contain an available carboxylic acid function whereby such analogs can be coupled to the amino-derivative of the label by the aforementioned peptide condensation reactions or bifunctional coupling agent reac-

tions (in the structural formulae below, n represents an integer, usually 1 through 6).

**Carbamazepine**

Dibenz[b,f]azepine is treated sequentially with phosgene, an  $\omega$ -aminoalkanol, and Jones reagent (chromium trioxide in sulfuric acid) according to the method of Singh, U.S. Pat. No. 4,058,511 to yield the following series of carboxylic acids:

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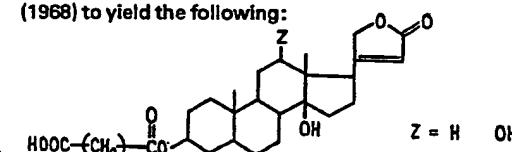
**Quinidine**

80 Following the method of Cook et al, *Pharmacologist* 17:218 (1975), quinidine is demethylated and treated with 5-bromovalerate followed by acid hydrolysis to yield a suitable carboxylic acid derivative.

**Digoxin and Digitalis**

The aglycone of the cardiac glycoside is treated with succinic anhydride and pyridine according to the method of Oliver et al, *J. Clin. Invest.* 47: 1035 (1968) to yield the following:

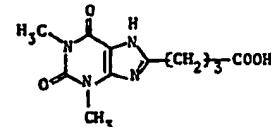
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**Theophylline**

Following the method of Cook et al, *Res. Comm. Chem. Path. Pharm.* 13:497 (1976), 4, 5-diamino-1, 3-dimethylpyrimidine-2, 6-dione is heated with 100 glutaric anhydride to yield the following:

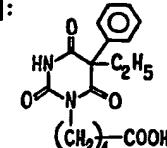
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**Phenobarbital and Primidone**

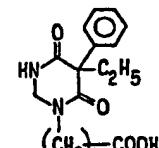
Sodium phenobarbital is heated with methyl 5-bromovalerate and the product hydrolyzed to the corresponding acid derivative of phenobarbital [Cook et al, *Quantitative Analytic Studies in Epilepsy*, ed. Kellaway and Peterson, Raven Press (New York 1976) pp. 39-58]:

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120 To obtain the acid derivative of primidone following the same Cook et al reference method, 2-thiophenobarbital is alkylated, hydrolyzed, and the product treated with Raney nickel to yield:

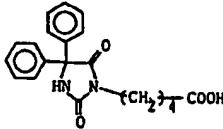
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**Diphenylhydantoin**

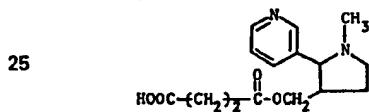
Following the method of Cook *et al*, *Res. Comm. Chem. Path. Pharm.* 5:767 (1973), sodium diphenylhydantoin is reacted with methyl 6-5 bromovalerate followed by acid hydrolysis to yield the following:

**Morphine**

Morphine free base is treated with sodium  $\beta$ -chloroacetate according to the method of Spector 16 *et al*, *Science* 168:1347 (1970) to yield a suitable carboxylic acid derivative.

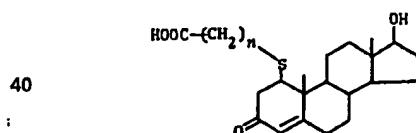
**Nicotine**

According to the method of Langone *et al*, 20 *Biochem.* 12(24):5025 (1973), *trans*-hydroxymethylnicotine and succinic anhydride are reacted to yield the following:

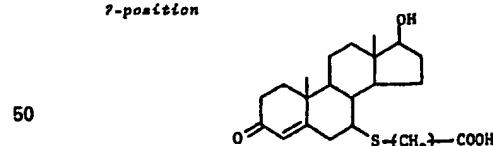
**Androgens**

Suitable carboxylic acid derivatives of testosterone and androstenedione linked through either the 1- or 7-position on the steroid nucleus are prepared according to the method of Bauminger *et al*, *J. Steroid Biochem.* 5:739 (1974). Following are representative testosterone derivatives:

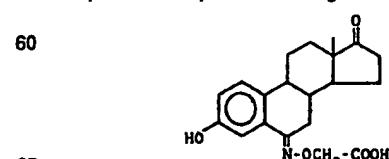
35      1-position



45      7-position

**Estrogens**

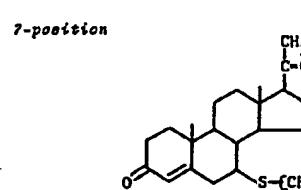
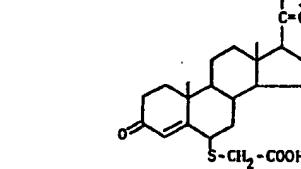
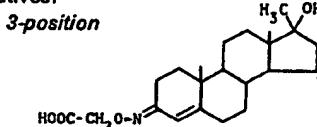
55      Suitable carboxylic acid derivatives of estrogens, e.g., estrone, estradiol and estriol, are prepared according to the method of Bauminger *et al*, *supra*, as represented by the following estrone derivative:



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**Progesterones**

Suitable carboxylic acid derivatives of progesterone and its metabolites linked through any of the 3-, 6- or 7-positions on the steroid nucleus are prepared according to the method of Bauminger *et al*, *supra*, as represented by the following progesterone derivatives:



The methods described above are but examples of the many known techniques for forming suitable carboxylic acid derivatives of haptens of analytical interest. The principal derivation techniques are discussed in *Clin. Chem.* 22:726 (1976) and include esterification of a primary alcohol with succinic anhydride [Abraham and Grover, *Principles of Competitive Protein-Binding Assays*, ed. Odell and Daughaday, J. B. Lippincott Co. (Philadelphia 1971) pp. 140-157], formation of an oxime from reaction of a ketone group with carboxymethyl hydroxylamine [*J. Biol. Chem.* 234:1090 (1959)], introduction of a carboxyl group into a phenolic residue using chloroacetate [*Science* 168:1347 (1970)], and coupling to diazotized *p*-aminobenzoic acid in the manner described in *J. Biol. Chem.* 235:1051 (1960).

The present invention will now be illustrated, but 115 is not intended to be limited, by the following Examples:

**Biotin Conjugate**

*A. Preparation of the Labeled Conjugate*  
120      The reaction sequence for this synthesis is described and shown schematically in *Anal. Chem.* 48:1933 (1976).

*4 - (3 - Chloro - 2 - hydroxypropylamino) - N - 125 methylphthalimide.*

Twenty-five grams (g) (0.142 mole) 4 - amino - N - methylphthalimide [Flitsch, *Chem. Ber.* 94:2494 (1961)] and 20.7 g (0.21 mole) 1 - chloro - 2, 3 - epoxypropane were added to 150 ml 2, 2, 2 - trifluoroethanol and the reaction mixture was heated to

reflux with stirring for 48 hours. Seventy to eighty ml of 2, 2, 2 - trifluoroethanol was removed by distillation and a heavy yellow precipitate formed when the remaining solution cooled to room temperature.

5 This precipitate was triturated with ethyl acetate, collected by filtration and dried to give 29.5 g (77% yield) of the desired phthalimide intermediate m.p. 136-138.5°C.

Analysis: Calculated for  $C_{12}H_{13}ClN_2O_3$ : C, 53.64; H, 4.88; N, 10.45  
Found: C, 53.87; H, 4.85; N, 10.81

4 - [3 - (*N* - Phthalimido) - 2 - hydroxypropylamino] - *N* - methylphthalimide.

15 The phthalimide intermediate prepared above (13.5 g, 0.05 mole) and 15.7 g (0.085 mole) potassium phthalimide were heated to reflux with stirring in 150 ml dimethylformamide for 24 hours. The dimethylformamide was removed and the residue was

20 washed with water and filtered. The yellow filter cake was recrystallized from acetic acid-water to give 12.8 g (67% yield) of the *bis*-phthalimide intermediate, m.p. 247-248.5°C.

Analysis: Calculated for  $C_{20}H_{17}N_3O_5$ : C, 63.32; H, 4.52; N, 11.08  
Found: C, 63.16; H, 4.38; N, 10.93

6 - (3 - Amino - 2 - hydroxypropylamino) - 2, 3 - dihydropthalazine - 1, 4 - dione.

30 The *bis*-phthalimide intermediate from above (5.0 g, 13.2 mmole), 90 ml absolute ethanol and 35 ml 95% hydrazine were refluxed with stirring for 4 hours. The solvent was removed under a vacuum and the resulting solid was dried for 24 hours under

35 vacuum at 120°C. This material was stirred for 1 hour with 70 ml of 0.1 N hydrochloric acid. The insoluble material was removed by filtration and the filtrate was adjusted to pH 6.5 with saturated sodium bicarbonate. The white precipitate which formed was collected by filtration and dried to give 2.2 g of the product (67% yield). After recrystallization from water, the compound decomposed at 273°C.

Analysis: Calculated for  $C_{11}H_{14}N_2O_3$ : C, 52.79; H, 5.64; N, 22.39  
45 Found: C, 52.73; H, 5.72; N, 22.64

The efficiency of the amino-derivative (i.e., the label derivative) in a chemiluminescent reaction and the detection limit of such derivative were determined as follows.

50 In determining efficiency, the label derivative and luminol (5 - amino - 2, 3 - dihydropthalazine - 1, 4 - dione) were oxidized individually at several levels in the picomolar range and related to the peak light intensities generated by a graph plot. Linear portions

55 of the resulting curves allowed calculation of change in peak light intensity per unit concentration for the label derivative and for luminol. Efficiency of the label derivative was expressed as a percentage of the slope produced with luminol.

60 Reaction mixtures (150  $\mu$ l) of the following composition were assembled in 6x50 mm test tubes mounted in a Dupont 760 Luminescence Biometer (E. I. duPont de Nemours and Co., Wilmington, Delaware USA) with a sensitivity setting of 820:50

65 mM sodium hydroxide, 0.07  $\mu$ M hematin (Sigma

Chemical Co., St. Louis, Missouri USA) and either the amino-derivative or luminol at varying concentrations in the picomolar (pM) range (diluted with H<sub>2</sub>O from a 1 mM stock solution in 0.1 M sodium carbonate, pH 10.5). Each mixture was incubated 10 minutes at room temperature and 10  $\mu$ l of 90 mM hydrogen peroxide was added to initiate the chemiluminescent reaction. Peak light intensity values were recorded from the instrument readings. All reactions were performed in triplicate and averaged. The efficiency of the label derivative was found to be 10%.

Detection limit was defined as the concentration of the label derivative that produced a peak light intensity one and a half times the background chemiluminescence in the reaction mixture. The detection limit for the label derivative was found to be 20 pM.

The N-ethylated derivative, 6 - [N - (3 - amino - 2 - hydroxypropyl) - N - ethylamino] - 2, 3 - dihydropthalazine - 1, 4 - dione, of the above-described amino-derivative was also prepared by treating 4 - amino - N - methylphthalimide with diethyl sulfate under reflux in 2, 2, 2 - trifluoroethanol and then following the same synthesis as described above to convert the N-ethylated intermediate through the phthalimide and *bis*-phthalimide intermediate stages to the N-ethylated amino-derivative. The efficiency of this compound in the hematin catalyzed chemiluminescent reaction was found to be 46% and the detection limit 5 pM.

6 - (3 - Biotinylamido - 2 - hydroxypropylamino) - 2, 3 - dihydropthalazine - 1, 4 - dione [biotin - isoluminol conjugate].

Biotin (0.29 g, 1.2 mmole) and 0.17 ml triethylamine were dissolved in 20 ml dry dimethylformamide under anhydrous conditions and cooled to -10°C. A solution of 0.141 ml ethyl chloroformate in 2.86 ml ether was added slowly and the reaction was stirred for 30 minutes. A precipitate which formed was separated by filtration. A suspension consisting of 600 mg (2.4 mmole) of the amino-derivative intermediate from above, 20 ml dry

105 dimethylformamide and 1 ml dry pyridine was added to the filtrate quickly. This mixture was stirred at -10°C for 30 minutes and then at room temperature overnight. During this period a solution was obtained. The dimethylformamide was removed by

110 distillation at 60°C and 0.10 mm Hg pressure. The oily residue was stirred with 50 ml of 0.1 N hydrochloric acid for 1 hour. A white solid which formed was filtered and washed with 0.1 N hydrochloric acid and then water. After drying under a vacuum at room

115 temperature overnight, 0.55 g (97% yield) of the labeled conjugate was obtained, m.p. 170-3°C.

Analysis: Calculated for  $C_{21}H_{28}N_4O_5S$ : C, 52.92; H, 5.92; N, 17.64  
Found: C, 51.69; H, 5.90; N, 17.63

**B. Binding Assays for Biotin and Avidin Using Enzyme Catalyzed Monitoring Reaction**

The chemiluminescent reaction system used in this example was based on the following reaction:



5

Nine specific binding reaction mixtures were prepared, each having a total volume of 140  $\mu\text{l}$  and each containing 0.1 M tris - (hydroxymethyl) - aminomethane hydrochloride buffer (Tris - HCl) at 10 pH 7.4 and biotin, biotin - isoluminol labeled conjugate (prepared as above), and avidin (added last) in the concentrations indicated in Table 1. After 5

minute incubation at 25°C, 10  $\mu\text{l}$  0.1 M Tris - HCl buffer at pH 7.4 containing 20 units/ml lactoperoxidase [Sigma Chemical Co., St. Louis, Missouri USA; assayed as described in *Methods in Enzymology* XVIIA, (1970) p. 653-Assay 2] was added to each reaction mixture. After incubation at 25°C for 2 additional minutes, 10  $\mu\text{l}$  of 0.95 mM hydrogen peroxide 20 in 10 mM Tris-HCl buffer at pH 7.4 was injected into each reaction mixture and the peak light intensity produced in each was measured using the Dupont Model 760 Bioluminescence Photometer. The results appear in Table 1.

TABLE 1

| reaction mixture | concentration of biotin ( $\mu\text{M}$ ) | concentration of biotin-isoluminol conjugate (nM) | concentration of avidin (units/ml) | peak light intensity |
|------------------|---|---|------------------------------------|----------------------|
| 1                | —   | —   | —                                  | 0.8                  |
| 2                | —   | —   | 0.14                               | 0.9                  |
| 3                | —   | 84  | —                                  | 1.9                  |
| 4                | —   | 84  | 0.14                               | 25.3                 |
| 5                | 4   | —   | —                                  | 0.8                  |
| 6                | 4   | 84  | —                                  | 2.2                  |
| 7                | 4   | —   | 0.14                               | 0.9                  |
| 8                | 4   | 84  | 0.14                               | 6.1                  |
| 9                | 1.3                                       | 84  | 0.14                               | 10.4                 |

25 Reactions 1, 2, 5 and 7 were controls and show that in the absence of biotin-isoluminol conjugate, only a low background amount of light was measured. The result of reactions 3 and 6 indicate that the biotin-isoluminol conjugate was active in the

mM hydrogen peroxide in 10 mM Tris-HCl buffer at pH 7.4 was injected into each reaction mixture and

30 chemiluminescent reaction and that the presence of free biotin had no significant effect on such activity. The result of reaction 4 shows that in the presence of

65 avidin, a binder for biotin, the activity of the biotin - isoluminol conjugate increased. This result is rather unexpected since one would anticipate that binding of avidin to the conjugate should limit the availability of the isoluminol moiety for the chemiluminescent reaction. The reason for the observed enhancement of light-production is not understood. A comparison

35 of the results of reactions 4, 8, and 9 demonstrate that the enhancement of light production is decreased inversely with the amount of free biotin present.

the peak light intensity produced in each was measured as in the previous experiment. The results appear in Table 2.

70

TABLE 2

| reaction mixture | concentration of biotin (nM) | peak light intensity |
|------------------|------------------------------|----------------------|
| 1                | 0                            | 23.5                 |
| 2                | 67                           | 21.1                 |
| 3                | 134                          | 15.5                 |
| 4                | 200                          | 12.6                 |
| 5                | 268                          | 12.3                 |
| 6                | 400                          | 8.1                  |

80 It was thus demonstrated that the magnitude of the peak light intensity produced by the chemiluminescent reaction system was an inverse function of the amount of biotin present in the specific binding reaction mixture. The present invention therefore provides labeled conjugates useful for determining the presence of ligands in a liquid medium.

#### 90 C. Binding Assay for Biotin Using Non-Enzymatic Monitoring Reaction

The chemiluminescent reaction system used in this example was based on the following reaction:



95 Sixteen specific binding reaction mixtures were prepared, each having a total volume of 150  $\mu\text{l}$  and each containing 0.1 M Tris-HCl at pH 8.0, 42 nM

45 and biotin can be determined using the present labeled conjugates and that according to the present invention the effect of binding between the labeling substance in the conjugate and a corresponding binding partner may be an enhancement, rather than inhibition, of the activity of the labeling substance.

50 A further experiment was conducted using the same lactoperoxidase-catalyzed monitoring reaction.

55 Six specific binding reaction mixtures were prepared, each having a total volume of 140  $\mu\text{l}$  and each containing 0.1 M Tris-HCl buffer at pH 7.4, 84 nM biotin-luminol conjugate (prepared as above), biotin at the concentrations indicated in Table 2, and 0.035 units/ml avidin (added last). After a 5 minute incubation at 25°C, 10  $\mu\text{l}$  of lactoperoxidase (20 units/ml) were added to each reaction mixture.

60 After an additional 2 minute incubation, 10  $\mu\text{l}$  0.95

biotin-isoluminol conjugate (prepared as above), biotin at the concentrations indicated in Table 3, and 0.12 units/ml avidin (added last). After incubation at 25°C for 5 minutes, 10 µl of dimethylformamide containing 0.15 M potassium superoxide ( $KO_2$ ) (Alpha Products, Beverly, Massachusetts USA) and 0.10 M 1, 4, 7, 10, 13, 16-hexaoxacyclooctadecane (Aldrich Chemical Co., Milwaukee, Wisconsin USA) were injected into each reaction mixture and the peak light intensity produced in each was measured as in the previous experiments. The results appear in Table 3.

TABLE 3

| 15 | reaction mixture | concentration of biotin (nM) | peak light intensity |
|----|------------------|------------------------------|----------------------|
| 1  |                  | 0                            | 38.5                 |
| 2  |                  | 13                           | 38.5                 |
| 3  |                  | 27                           | 34.3                 |
| 20 | 4                | 40                           | 36.1                 |
|    | 5                | 53                           | 35.2                 |
|    | 6                | 67                           | 36.2                 |
|    | 7                | 101                          | 34.0                 |
|    | 8                | 133                          | 31.7                 |
| 25 | 9                | 166                          | 29.1                 |
|    | 10               | 200                          | 24.2                 |
|    | 11               | 267                          | 22.8                 |
|    | 12               | 333                          | 20.5                 |
|    | 13               | 400                          | 13.4                 |
| 30 | 14               | 534                          | 8.6                  |
|    | 15               | 667                          | 8.3                  |
|    | 16               | 800                          | 7.0                  |

It was demonstrated that the magnitude of the peak light intensity produced by the chemiluminescent reaction system was an inverse function of the amount of biotin present in the specific binding reaction mixture. The present invention therefore provides labeled conjugates useful for determining the presence of ligands in a liquid medium.

#### Thyroxine Conjugates

##### A. Preparation of Labeled Conjugates

Following are descriptions of the preparation of the labeled thyroxine conjugates 6 - N - [2 - hydroxy - 3 - (thyroxinylamido) propyl] amino - 2, 3 - dihydropthalazine - 1, 4 - dione and 6 - {N - ethyl - N - [2 - hydroxy - 3 - (thyroxinylamido) propyl] amino} - 2, 3 - dihydropthalazine - 1, 4 - dione. The reaction sequences for these syntheses are outlined in Tables 4 and 5.

##### N - Trifluoroacetylthyroxine (2).

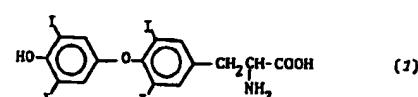
A solution of 20 grams (g) [25.6 millimole (mmol)] of L-thyroxine (1) (Sigma Chemical Co., St. Louis, Missouri USA) in 240 milliliters (ml) of ethyl acetate containing 46 ml of trifluoroacetic acid and 7.6 ml of trifluoroacetic anhydride was stirred at 0°C for one hour. Upon adding 200 ml of water ( $H_2O$ ), a suspension formed that was saturated with sodium chloride. The organic phase was separated, washed with saturated aqueous sodium chloride solution, dried over anhydrous magnesium sulfate, filtered and evaporated. When dry, the crystalline residue amounted to 21.3 g of the N-protected thyroxine

derivative (2). A sample was recrystallized from ether-pentane to give fine white crystals, melting point (m.p.) 233-235°C (decomposed).

Analysis: Calculated for  $C_{17}H_{10}F_3I_4NO_5$ : C, 23.39; H, 1.15; N 1.60  
Found: C, 23.23; H, 1.12; N, 1.59  
Infrared Spectrum (KCl): 1700  $\text{cm}^{-1}$  (carbonyl)  
Optical Rotation  $[\alpha]_D^{25} = -14.97^\circ$  (c 1.0, dimethylsulfoxide)

75

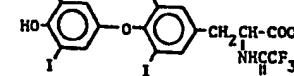
TABLE 4



80



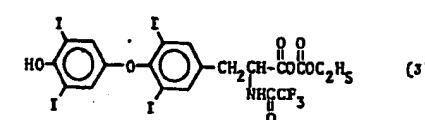
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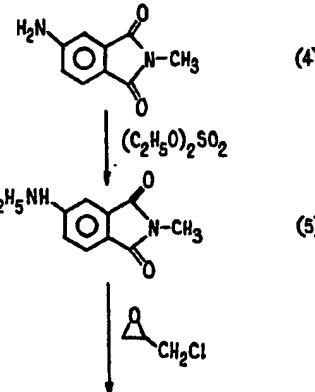
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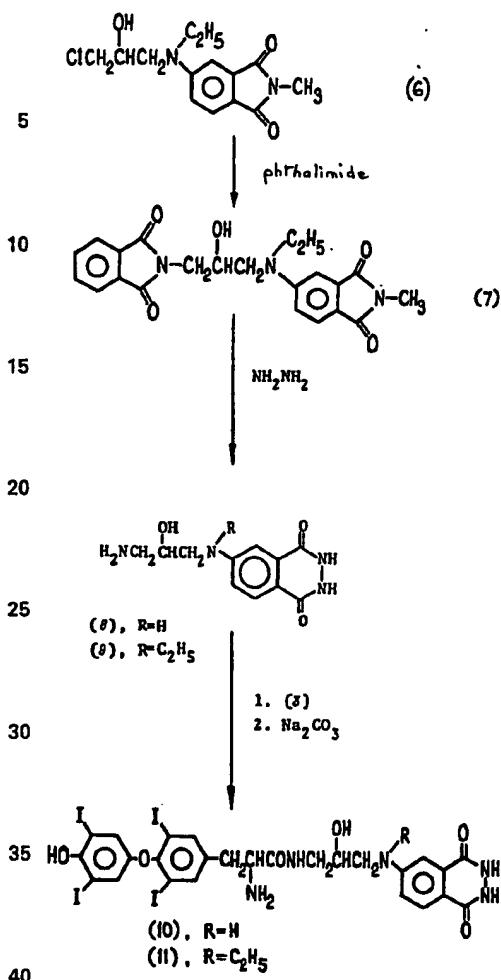
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125

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TABLE 5





*N*-Trifluoroacetylthyroxinyl Ethyl Carbonic Anhydride (3).

A mixture of 0.17 ml of triethylamine and 1.05 g (1.2 mmol) of N-trifluoroacetylthyroxine (2) was dissolved in 20 ml of dry dimethylformamide at -10°C under anhydrous conditions [Knappe et al, Biochem. Z. 338:599 (1963)]. To this was added a solution of 0.14 ml (1.2 mmol) of ethyl chloroformate in 2.9 ml of dry ether. After 30 minutes the precipitate of triethylammonium chloride was removed by filtration. The filtrate, now containing the anhydride (3), was used without isolation in the reaction described below to form labeled conjugate (10).

55 4-N-Ethylamino-N-methylphthalimide (5).

A mixture of 10 g (0.057 mol) of 4-amino-N-methylphthalimide (4) [Flitsch, Chem. Ber. 94:2494 (1961)], 17.5 g (0.11 mol) of diethyl sulfate, and 100 ml of 2,2,2-trifluoroethanol was refluxed for one day. The reaction mixture was cooled, concentrated under reduced pressure, and the residue partitioned between 250 ml of ethyl acetate and 100 ml of saturated sodium bicarbonate solution containing 10 ml of triethylamine. The ethyl acetate phase was separated, washed with saturated aqueous sodium

chloride solution, dried over anhydrous magnesium sulfate, filtered, and evaporated. The residue was recrystallized twice from acetone-hexane and then from aqueous methanol to give 3.4 g (29% yield) of the phthalimide (5) as fine yellow crystals, m.p. 157°C.

Analysis: Calculated for C<sub>11</sub>H<sub>12</sub>N<sub>2</sub>O<sub>2</sub>: C, 64.69; H, 5.92; N, 13.71;

Found: C, 64.00; H, 5.71; N, 13.37

75 NMR Spectrum (C<sub>6</sub>D<sub>6</sub>N): δ 0.5 (t, J = 7 Hz, 3H), 2.1 (s, 3H)

4-[N-(3-Chloro-2-hydroxypropyl)-N-ethylamino]-N-methylphthalimide (6).

80 A mixture of 3.1 g (15 mmol) of the phthalimide (5), 1.2 g (15 mmol) of 1-chloro-2,3-epoxyp propane, and 30 ml of 2,2,2-trifluoroethanol was refluxed for 24 hours. At the end of this time, another 1.2 g of 1-chloro-2,3-epoxypropane was added.

85 After heating for an additional 24 hours, the reaction was cooled and evaporated and the residue chromatographed on 150 g of silica gel 60 (E. Merck, Darmstadt, West Germany), eluting with a 9:1 mixture (v:v) of carbon tetrachloride and acetone. Fifteen ml fractions were collected. Fractions numbered 90 to 160 were combined and evaporated to give 3 g of a yellow-red oil. The oil was crystallized from acetone-hexane and recrystallized twice from aqueous methanol to give 1 g (22% yield) of the phthalimide (6) as fine yellow needles, m.p. 123°C.

Analysis: Calculated for C<sub>14</sub>H<sub>17</sub>CIN<sub>2</sub>O<sub>3</sub>: C, 56.66; H, 5.78; N, 9.44

Found: C, 56.50; H, 5.93; N, 9.26

100 NMR Spectrum (C<sub>6</sub>D<sub>6</sub>N): δ 0.65 (t, 3H, J = 8 Hz), 2.6 (s, 3H)

4-[N-Ethyl-N-[2-hydroxy-3-(N-phthalimido)propyl]amino]-N-methylphthalimide (7).

105 A mixture of 25 g (0.08 mol) of the phthalimide (6), 23 g (0.13 mol) of potassium phthalimide, and 150 ml of dry dimethylformamide was refluxed for 36 hours. Removal of the solvent left a brown residue that was triturated with methanol to give 19 g of yellow solid. Recrystallization from aqueous acetic acid, then

110 from aqueous methanol, gave 16 g (49% yield) of the bis-phthalimide (7) as a yellow solid, m.p. 158-160°C.

Analysis: Calculated for C<sub>22</sub>H<sub>23</sub>N<sub>3</sub>O<sub>6</sub>: C, 64.85; H, 5.20; N, 10.31

Found: C, 64.81; H, 4.97; N, 10.54

115 NMR Spectrum (d<sub>6</sub>-DMSO): δ 1.2 (t, 3H, J = 6 Hz), 3.0 (s, 3H)

6-N-(3-Amino-2-hydroxypropyl)amino-2,3-dihydrophthalazine-1,4-dione (8)

120 This compound was prepared according to the method described both above relating to the biotin conjugate and in Anal. Chem. 48:1933 (1976). As reported above, the efficiency of this amino-derivative (8) in the hematin catalyzed chemiluminescent reaction was 10% and the detection limit 20 pM.

6-[N-(3-Amino-2-hydroxypropyl)-N-ethylamino]-2,3-dihydrophthalazine-1,4-dione (9)

130 A mixture of 15 g (0.037 mol) of the bis-

phthalimide (7), 60 ml of 95% hydrazine, and 300 ml of absolute ethanol was refluxed for 3 hours. The reaction was cooled, evaporated to dryness, and the crystalline residue dried at 40°C/0.05 mm Hg over 6 night. The residue was then dried at 120°C/0.05 mm Hg for 4 hours. The resulting solid was stirred for 3 hours in dilute hydrochloric acid and filtered. When the pH of the filtrate was adjusted to 7.0, a precipitate formed amounting to 4.6 g (46% yield) of the 10 amino-phthalhydrazide (9), m.p. 207-210°C (decomposed). A small sample was recrystallized from H<sub>2</sub>O to give white crystals, m.p. 208-211°C (decomposed).

Analysis: Calculated for C<sub>13</sub>H<sub>16</sub>N<sub>4</sub>O<sub>3</sub>: C, 56.10; H, 6.52; N, 20.13

15 Found: C, 55.81; H, 6.50; N, 20.35

The efficiency of the amino-derivative (9) and its detection limit were determined in the same manner as described above for the amino-derivative (8). The efficiency was found to be 46% and the detection 20 limit 5 pM.

**6 - N-[2 - Hydroxy - 3 - (thyroxinylamido) propyl] amino - 2, 3 - dihydropthalazine - 1, 4 - dione (10).**

A suspension of 600 mg (2.4 mmol) of the amino-25 derivative (8) in 20 ml of dry dimethylformamide containing 1 ml of pyridine was stirred under argon for one hour. It was then drawn up into a syringe and added all at once to a -10°C solution of 1.2 mmoles of N - trifluoroacetylthyroxinyl ethyl carbonic anhydride (3) in 20 ml of dimethylformamide. After stirring for 20 minutes at -10°C, the reaction was allowed to warm to room temperature and stirred overnight. Solvent was removed under high vacuum. The solid residue was stirred for 40 minutes in dilute hydroch-35 loric acid, then filtered and dried under high vacuum to give 1.27 g of a free-flowing powder.

The trifluoroacetyl protecting group was removed by stirring 1.0 g of this powder for 5 hours in a methanol/H<sub>2</sub>O solution. The pH of the solution was 40 adjusted to 10.7 with solid sodium carbonate. The pH was reduced to 7.0 with hydrochloric acid and a white precipitate collected and dried. When dry this amounted to 700 mg (59% yield) of the labeled conjugate (10) as a yellowish-white powder, m.p. 235°C 45 (decomposed).

Analysis: Calculated for C<sub>28</sub>H<sub>23</sub>I<sub>4</sub>N<sub>6</sub>O<sub>8</sub>: C, 30.95; H, 2.30; N, 6.94

Found: C, 30.17; H, 2.33; N, 6.33

**50 6 - {N - Ethyl - N - [2 - hydroxy - 3 - (thyroxinylamido) propyl] amino} - 2, 3 - dihydropthalazine - 1, 4 - dione (11).**

A solution of 1.06 g (1.2 mmol) of N - trifluoro - acetylthyroxine (2) in 20 ml of dimethylformamide 55 containing 0.17 ml of triethylamine was cooled to -10°C. To this was added 0.14 ml (1.2 mmol) of ethyl chloroformate. After 30 minutes at this temperature the solution, now containing the mixed anhydride (3), was filtered to remove precipitated triethylamine 60 hydrochloride and added to a suspension of 668 mg (2.4 mmol) of the amino-derivative (9) in 20 ml of dimethylformamide. After stirring two days at room temperature, the solvent was removed under vacuum, and the residue washed with 10% hydrochloric 65 acid, collected by filtration, and dried.

To remove the trifluoroacetyl blocking group, the solid was dissolved in 50 ml of 0.1 M sodium carbonate (pH 10.5) to which was added a small amount of dimethylformamide to achieve solution. After one 70 day at room temperature, it was evaporated to dryness. The residue was taken up in 30 ml of H<sub>2</sub>O, and the pH adjusted to 7.2 with dilute hydrochloric acid. A solid precipitated that was collected and dried at 60°C under high vacuum to give 600 mg (50% yield) 75 of the labeled conjugate (11) as white crystals, m.p. >240°C (decomposed).

Analysis: Calculated for C<sub>28</sub>H<sub>27</sub>I<sub>4</sub>N<sub>6</sub>O<sub>8</sub>: C, 32.42; H, 2.62; N, 6.75

Found: C, 29.81; H, 2.69; N, 5.45

**80 B. Binding Assay for Thyroxine**

Competitive binding reaction mixtures (120 µl) were assembled in triplicate by combining the following reagents: 12 µl of 100 mM Tris-HCl (pH 8.8), 12 µl of 77 nM labeled conjugate (11) [labeled conjugate (10) could be used as well] in 10 mM Tris-HCl (pH 8.8), varying volumes of 40 nM thyroxine in the same buffer, 10 µl of a preparation of antibody to thyroxine in 5 mM phosphate buffer (pH 6.7), and a sufficient volume of H<sub>2</sub>O to make a final volume of 90 120 µl. After a 1 hour incubation at room temperature, the free- and bound-species of the labeled conjugate were separated for each reaction mixture by applying a 100 µl aliquot to small columns (0.3 µl bed volume) of Sephadex G-25 (Pharmacia Fine 95 Chemicals, Uppsala, Sweden) previously washed with 10 mM Tris-HCl (pH 8.8). The bound-species of the labeled conjugate was eluted from the column with 0.5 ml of the same buffer leaving the free-species in the column.

100 An aliquot (115 µl) of each column effluent was added to 35 µl of 0.29 µM hematin (Sigma Chemical Co., St. Louis, Missouri USA) and 214 mM sodium hydroxide in a 6x50 mm test tube. Each tube was placed in the Dupont 760 Biometer and 10 µl of 90 105 mM hydrogen peroxide in 10 mM Tris-HCl (pH 7.4) were added. The resulting peak intensity of the light produced in each chemiluminescent reaction was recorded from the instrument reading and the results from the triplicate runs were averaged.

110 The relationship of the amount of thyroxine in the binding reaction to peak light intensity is shown in Table 6 below.

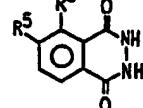
TABLE 6

|  | volume of thyroxine solution added (µl) | peak light intensity |
|--|---|----------------------|
|  | 0                                       | 13.8                 |
|  | 12                                      | 13.5                 |
|  | 48                                      | 3.5                  |

120 The results demonstrate that the labeled conjugate of the present invention is useful in binding assays for determining a ligand in a liquid medium.

**CLAIMS**

125 1. A compound of the formula:



130

wherein one of R<sup>a</sup> and R<sup>b</sup> is hydrogen and the other is -NR<sup>c</sup>R<sup>d</sup>; R<sup>e</sup> is hydrogen or straight chain alkyl containing 1-4 carbon atoms and R<sup>f</sup> is



wherein  $n = 1$  to  $3$ .

10 2. A compound according to claim 1 wherein R<sup>a</sup> is -NR'R".  
3. A compound according to claim 1 or 2, wherein n = 1.  
4. A compound according to any of claims 1 to 3, wherein R' is hydrogen.

15 5. A compound according to any of claims 1 to 3, wherein R' is ethyl.  
6. 6-(3-Amino-2-hydroxypropylamino)-2,3-dihydrophthalazine-1,4-dione.

20 7. 6-[N-(3-Amino-2-hydroxypropyl)-N-ethyl-amino]-2,3-dihydrophthalazine-1,4-dione.

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